

1 2 FIG S1. One-dimensional projection of NanoString data distinguishes susceptible 3 from resistant isolates. Panels show one-dimensional projections (squared projected 4 distance, SPD (1, 2)) of NanoString heatmap data from Fig. 1 for clinical isolates of E. 5 coli (top panels) and K. pneumoniae (bottom panels) treated across (a) fluoroquinolones, 6 (b) aminoglycosides, and (c) beta-lactams, binned by CLSI classifications (S, susceptible; 7 I, intermediate; R, resistant). By definition, an SPD of 0 indicates a transcriptional 8 response to antibiotic equivalent to that of an average susceptible strain, while an SPD 9 of 1 indicates a response equivalent to that of an average resistant strain. Data are 10 summarized as box-and-whisker plots in the style of Tukey (3), where boxes extend from the 25<sup>th</sup> to 75<sup>th</sup> percentile for each category, with a line at the median, and whiskers extend 11 12 from the minimum to the maximum.



13

FIG S2. GoPhAST-R detects carbapenemase (CPase) and extended-spectrum beta-14 lactamase (ESBL) gene content, augmenting phenotypic AST. Top panels show 15 16 GoPhAST-R detection of select CPase and ESBL transcript content in (a) *E. coli* and (b) 17 K. pneumoniae strains selected for BL treatment. Heatmap intensity reflects normalized, 18 background-subtracted, log-transformed NanoString data from probes for the indicated 19 gene families, as described (1). Color scales indicate range of log<sub>2</sub>[counts – background] 20 for these genes in the respective heatmap(s). Bottom panels show the GoPhAST-R 21 phenotypic AST heatmaps for ertapenem treatment from Fig. 1c. \*Note strain 14 displays phenotypic ertapenem resistance without detectable CPase or ESBL from our panel. 22 23

DATASET S1. Clinical isolates included in this study. Table shows the strains used
in this study with corresponding isolate source, relevant MICs, relevant resistance
mechanisms found by WGS analysis with ResFinder, and NCBI BioSample number.
Genotypic resistance determinants are color-coded by three general mechanisms of
resistance: permeability alterations (blue), target site modifications (orange), and drug
modifications (purple).

31		References
32 33	1.	Bhattacharyya RP, Bandyopadhyay N, Ma P, Son SS, Liu J, He LL, Wu L,
34		Khafizov R, Boykin R, Cerqueira GC, Pironti A, Rudy RF, Patel MM, Yang R,
35		Skerry J, Nazarian E, Musser KA, Taylor J, Pierce VM, Earl AM, Cosimi LA,
36		Shoresh N, Beechem J, Livny J, Hung DT. 2019. Simultaneous detection of
37		genotype and phenotype enables rapid and accurate antibiotic susceptibility
38		determination. Nat Med 25:1858-1864.
39	2.	Barczak AK, Gomez JE, Kaufmann BB, Hinson ER, Cosimi L, Borowsky ML,
40		Onderdonk AB, Stanley SA, Kaur D, Bryant KF, Knipe DM, Sloutsky A, Hung DT.
41		2012. RNA signatures allow rapid identification of pathogens and antibiotic
42		susceptibilities. Proc Natl Acad Sci U S A 109:6217-6222.
43	3.	McGill R, Tukey JW, Larsen WA. 1978. Variations of Box Plots. The American
44		Statistician 32:12-16.